

A Simple, Quick & Low Cost Semi-Automated Clean-Up and Concentration Solution for the Analysis of 209 PCB Congeners and other POPs

Fluid Management Systems
Watertown MA



Introduction (1)

- **POPs (PCDD/Fs, PCBs) continue to attract interest around the world due to strict regulations enforced in many countries**
- **Rapid and quality sample clean up and analysis is needed for many laboratories processing samples**
- **Processing times and cost are important considerations**
- **In the US, EPA methods SW-846, 1613, 1668 and 8082A are used for PCBs and PCDD/Fs work**

Introduction (2)

- **Sample extracts in DCM, hexane or toluene**
- **Cleanup for analysis of all 209 PCBs in common in North America - extracts are often in toluene after Soxhlet Extraction**
- **Dioxins and furans can also be run**
- **Extractable Petroleum Hydrocarbons are analyzed in soils for aliphatics and aromatics (PAHs)**
- **Individual states in US have methods available**

Challenges of POPs Sample Prep

- Labor intensive, prone to error
- Compliance with regulatory procedures and accreditation (lengthy method validation)
- Strict QA/QC requirements
- Sample matrix complexity
- Native background and interferences (can be orders of magnitude higher than analytes)
- Pico/femto-gram analyses require ultra pure extract and excellent instrument sensitivity

Automated Sample Prep

- **Advantages of Automated Sample Prep**

- Rapid Turn Around Time: 30 to 45 Minutes for 6 Samples
- Cleaner Background Interferences: Closed Loop System
- Quality Results: Certified Pre-packaged Columns
- Green Technology: Lower solvent and power use
- QA/QC & Accreditation Requirements: Easier to Manage
- Computerized Method: Instrumentation based prep



Manual Sample Prep

- **Advantages of Manual Sample Prep**
 - Most labs use a Manual Methods for the following reasons:
 - No electronics or mechanical components to fail
 - No down time due to the system failure
 - No service contract
 - No capital equipment cost



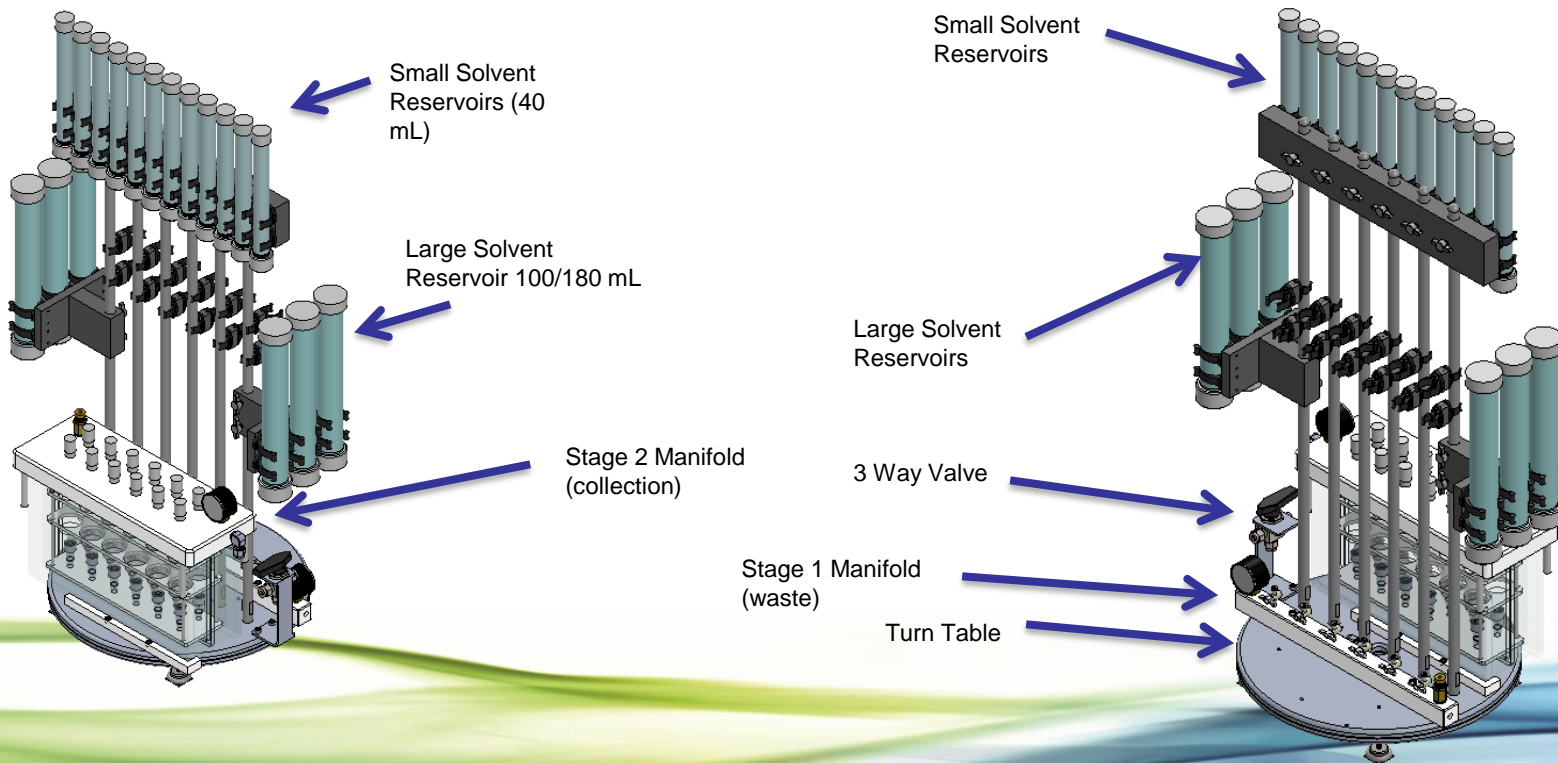
Semi-Automated System

Specification:

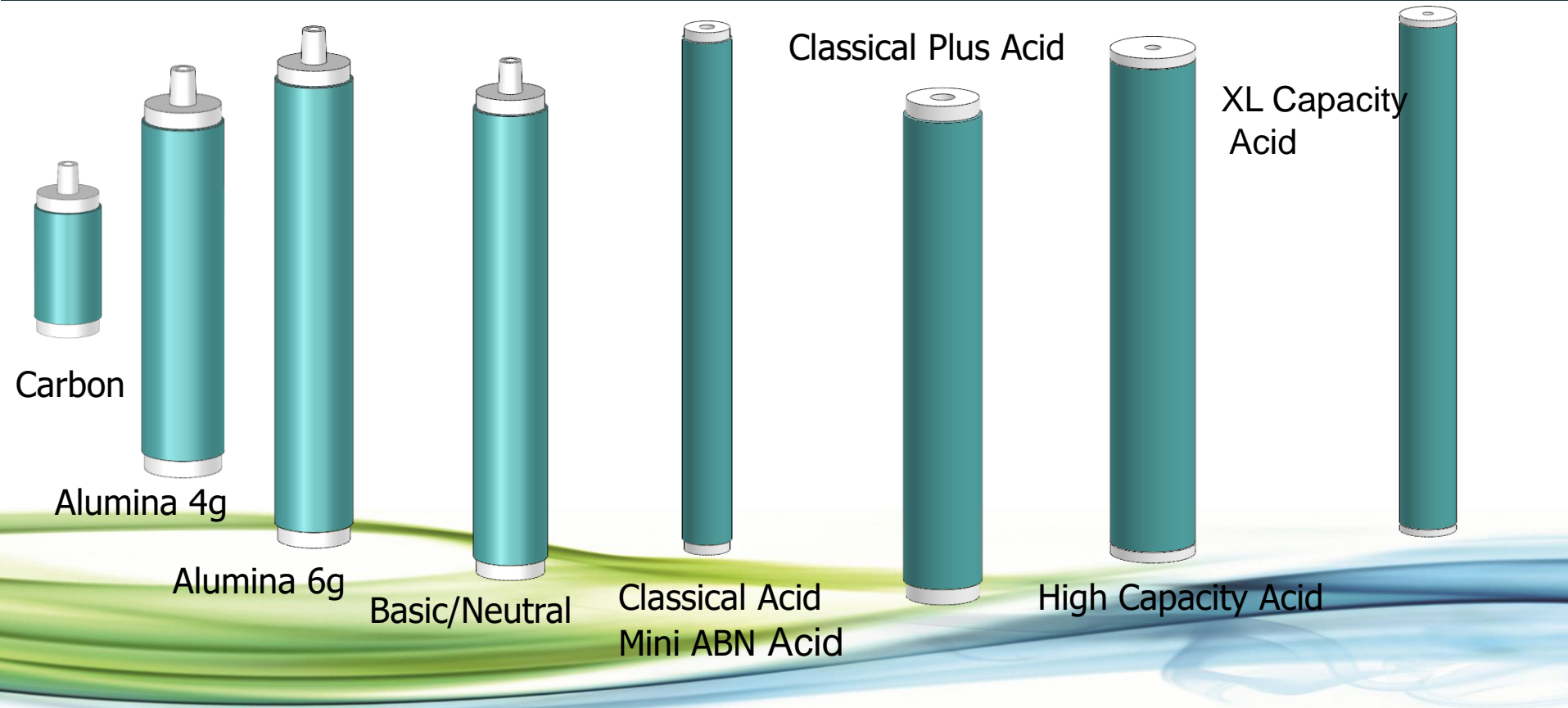
- Simple to run, no computerized instrumentation
- Fast: 30 to 45 min
- Closed loop system to give a clean background, low level detection
- Use certified pre-packaged columns
- Green technology, only vacuum pump uses power
- Low solvents, as low as 160 mL for serum
- Economical column kits, choice of low fat and high fat column kits
- No capital equipment cost
- No electronics or mechanical equipment to fail
- No downtime



Characteristics of Semi-Automated System (EZPrep)

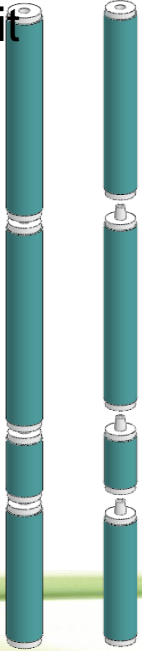


Columns (1)

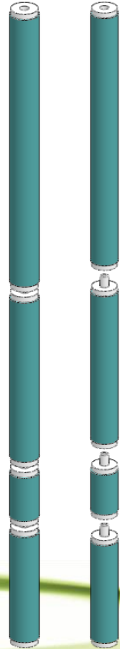


Columns (2)

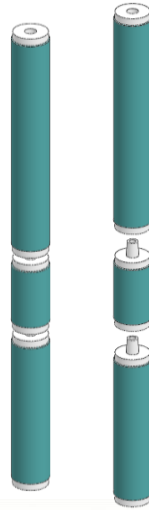
Classical
Kit



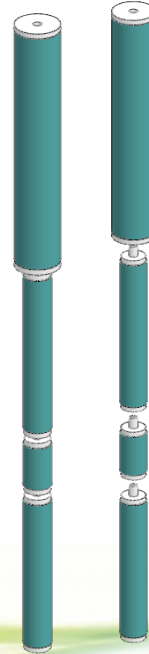
Classical Plus
Kit



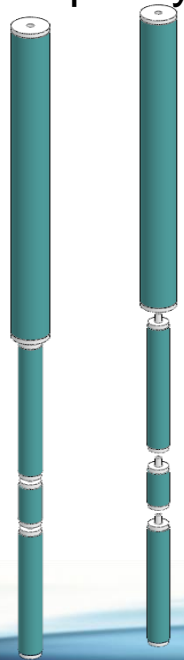
Mini Kit



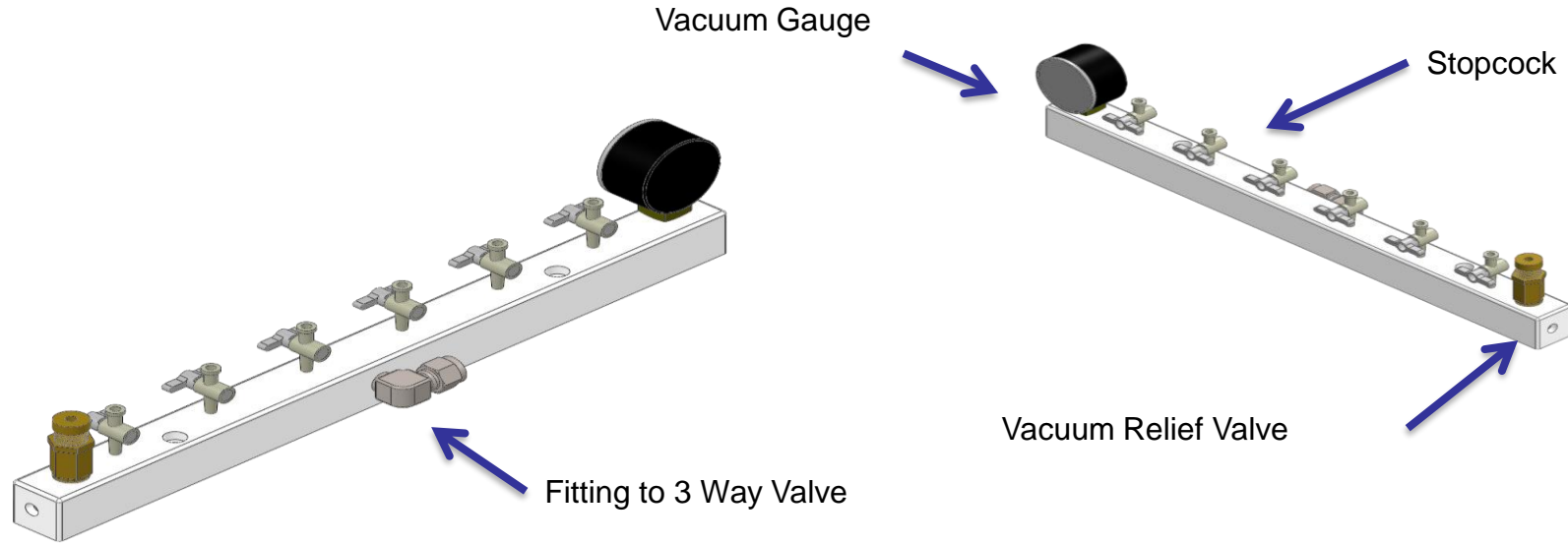
High Capacity
Kit



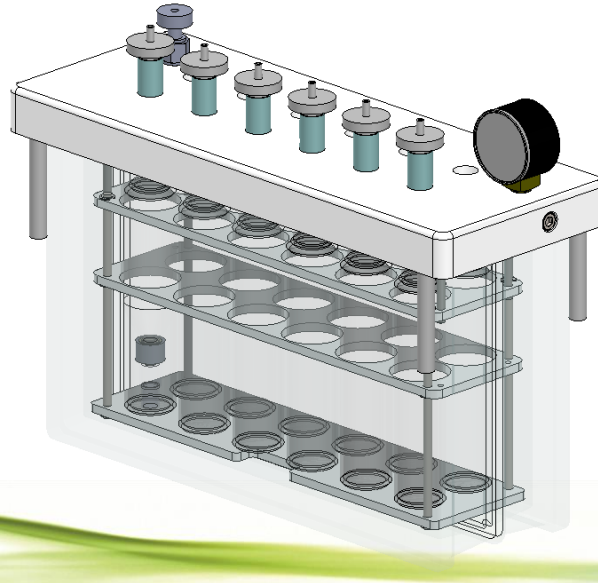
XL Capacity
Kit



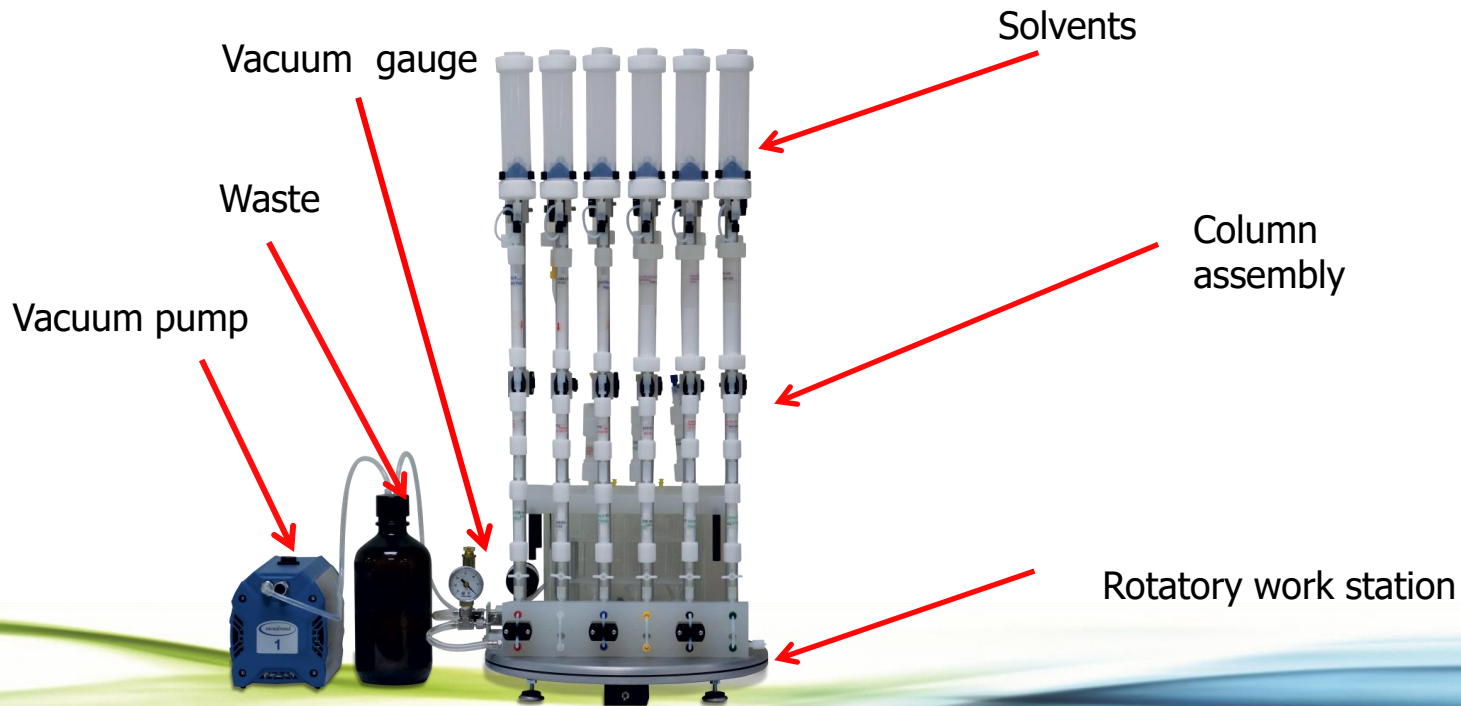
Stage 1 Manifold



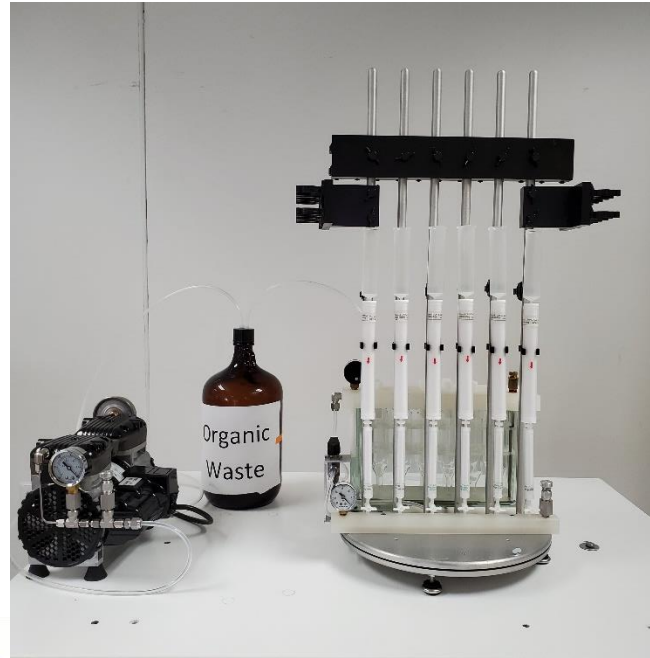
Stage 2 Manifold



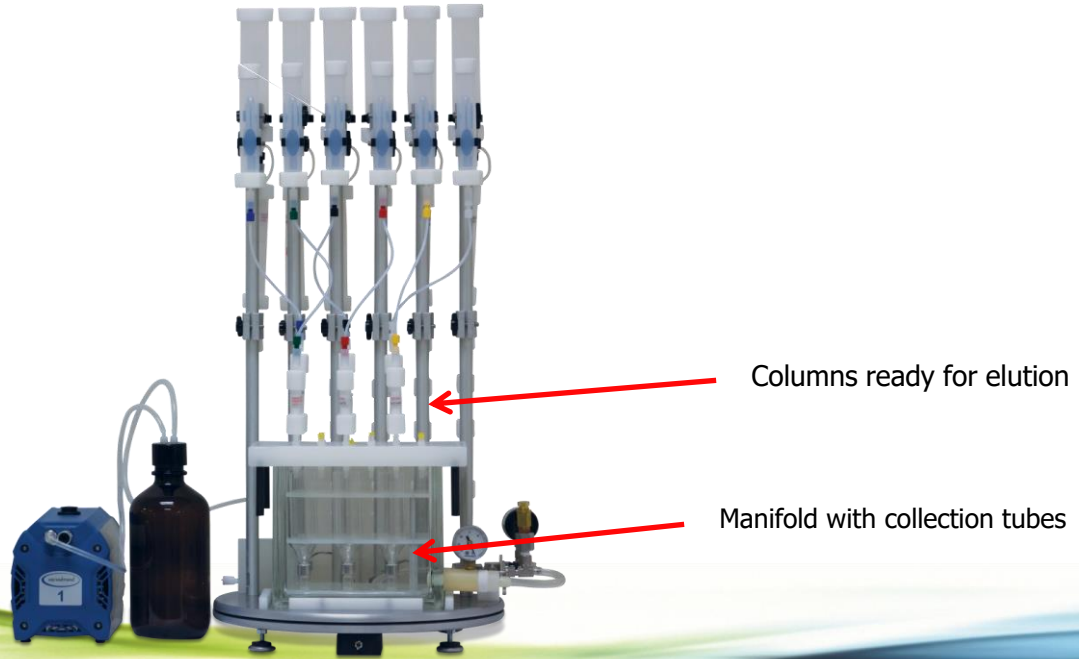
Stage 1: to waste



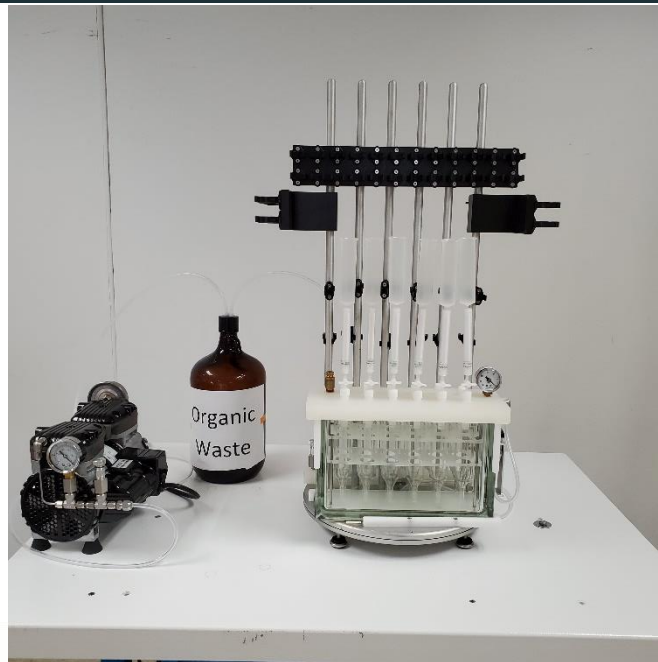
EZPrep Stage 1



Stage 2: collect



EZPrep Stage 2 Fractions



Attributes

- **Closed loop system:**
 - **Eliminates background contaminants**
 - **No washing needed.**
 - **Capped solvent reservoirs**
- **Optimized for solvent reduction while obtaining highest possible recoveries**
- **Easy sample loading on top of silica column via injection or syringe vial**
- **Columns connect easy with SNAP connections**



Extracts in hexane - PCBs

- **Stage 1: Connect High Capacity Acid Silica and Alumina (no Carbon) and condition with 60 mL of hexane (vacuum, waste)**
- **Stage 2: Load sample (in hexane, collect Fraction # 1), rinse loading vials with hexane, elute with 160 mL hexane (collect Fraction # 1), remove acid silica, elute alumina with 50 mL dichloromethane (collect Fraction # 1)**
- **All 209 PCBs are now in Fraction # 1**

SuperVap 6 Concentrator 250 mLs



SuperVap Concentration/Evaporation

- **System pre-heated to 55-60 °C.**
- **Samples evaporated at stable T under 5-6 psi nitrogen.**
- **1 mL extract vial transferred to GC vial (can have direct-to-vial feature).**
- **Recovery standards added (nonane/dodecane).**
- **Extract taken to 10 uL volume with a gentle stream of nitrogen at ambient temperature.**



SuperVap 24 position GC vial Concentrator



Direct-to-Vial

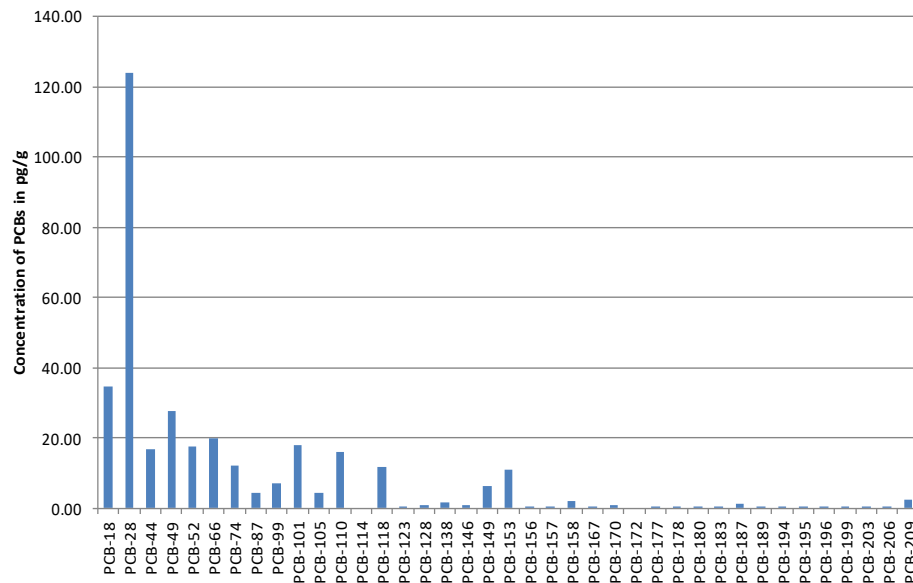


GC vial

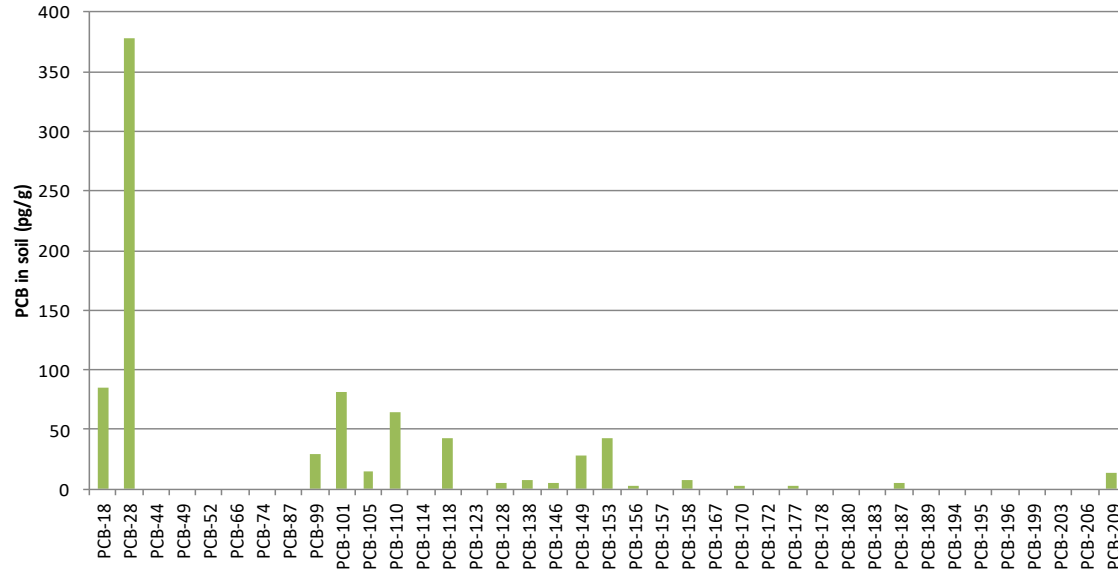
DFS HRGC/HRMS



Native PCBs in Serum extract



Native PCBs in Soil extract

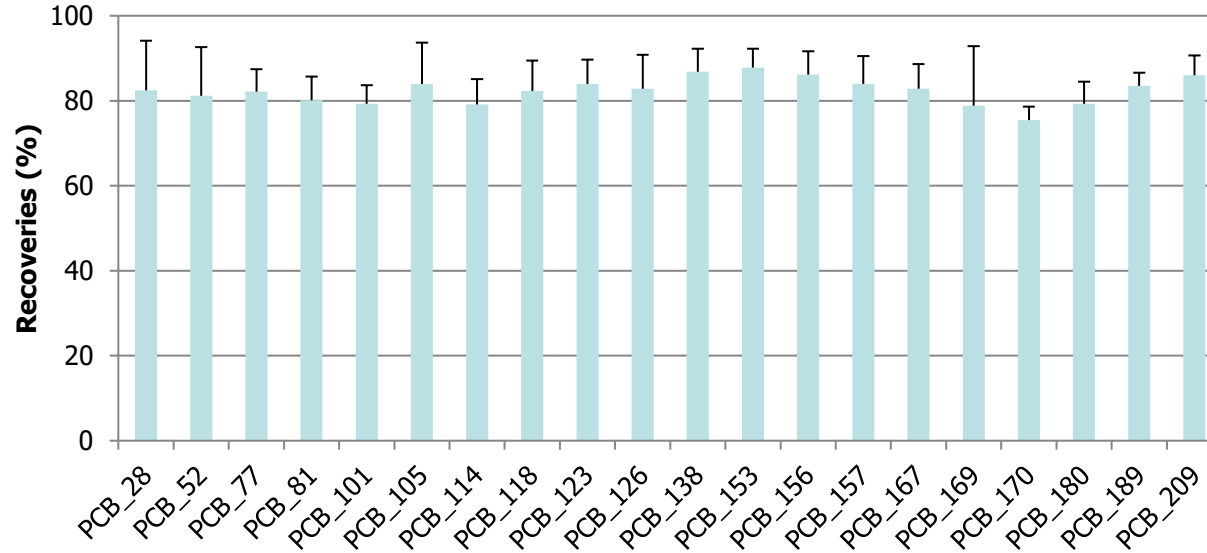


Extracts in toluene -PCBs

- Stage 1: Connect High Capacity Acid Silica and Alumina (no Carbon) and condition with 60 mL of hexane (vacuum, waste)
- Stage 2: Load sample (in 2-10 mL toluene, collect Fraction # 1), rinse loading vials with hexane, elute with 60 mL hexane (collect Fraction # 1), remove acid silica, elute alumina with 50 mL dichloromethane (collect Fraction # 1)
- All 209 PCBs are now in Fraction # 1

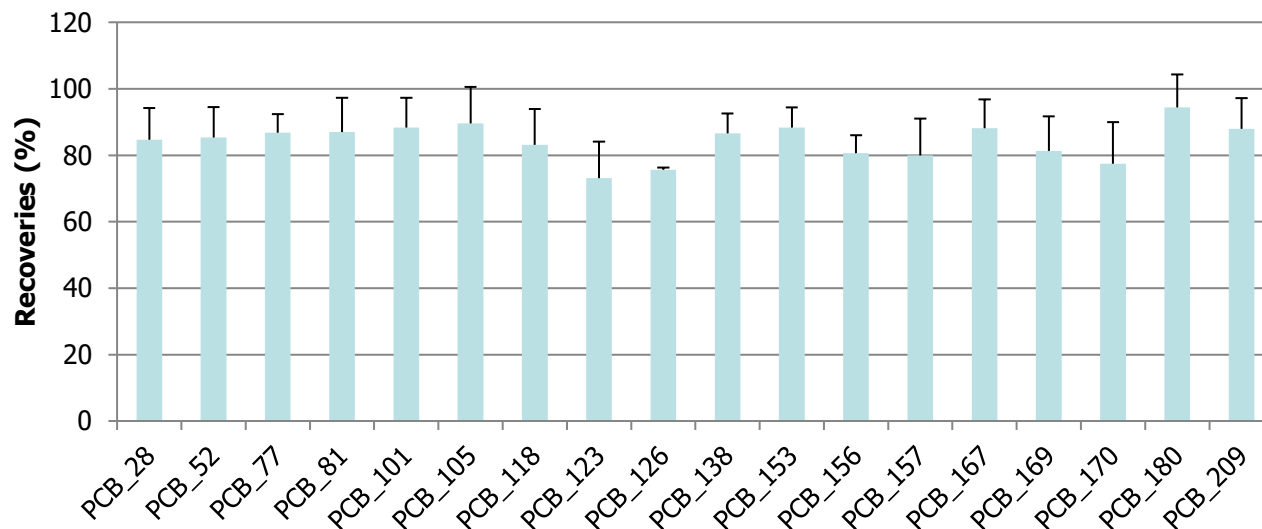


13C recoveries PCBs soil



10 g soil in
toluene, n=6

^{13}C recoveries PCBs salmon

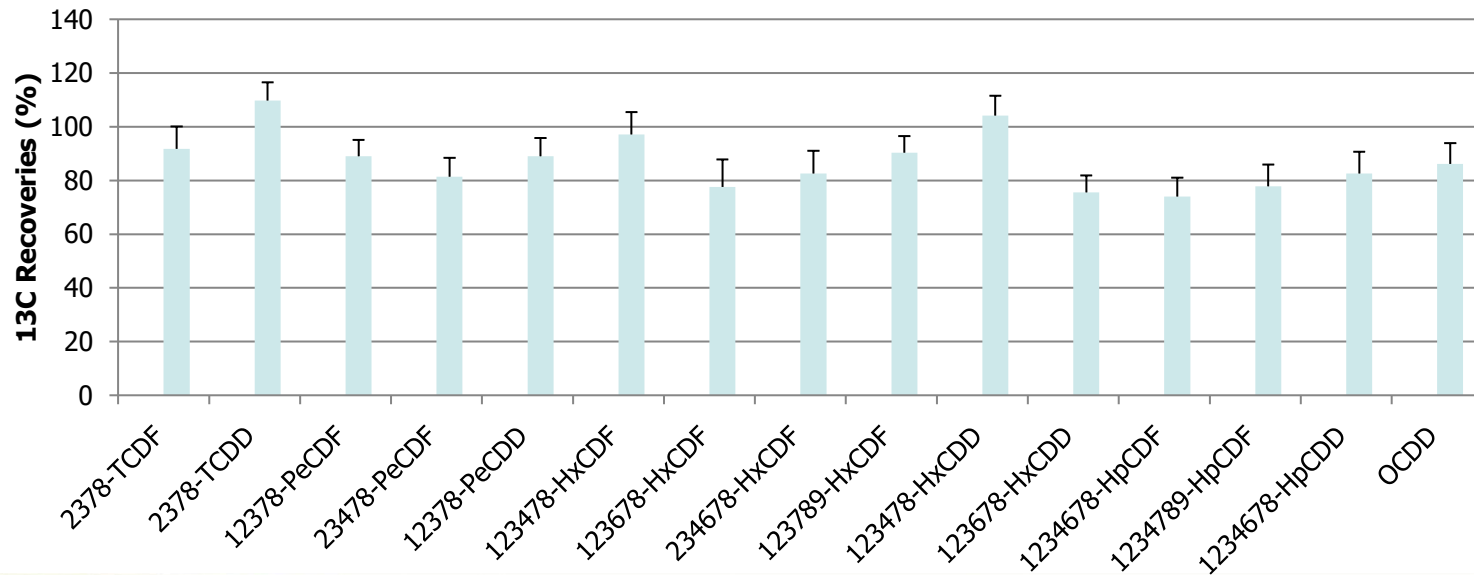


2 g salmon in
toluene, n=6

Extracts in hexane - PCDD/Fs

- **Stage 1: Connect High Capacity Acid Silica and Alumina (no Carbon) and condition with 60 mL of hexane (vacuum, waste)**
- **Stage 2: Load sample (in hexane, collect Fraction # 1), rinse loading vials with hexane, elute with 160 mL hexane (collect Fraction # 1), remove acid silica, elute alumina with 30 mL 10% dichloromethane in hexane (collect Fraction # 1)**
- **All 209 PCBs are now in Fraction # 1**
- **Stage 1 Attach carbon column to alumina, elute alumina-carbon with 50 mL dichloromethane (vacuum, waste), all PCDD/Fs now on carbon**
- **Stage 2 Discard alumina, elute carbon in reverse with 50 mL toluene, collect Fraction # 2 with all PCDD/Fs**

13C Recoveries PCDD/Fs - Water

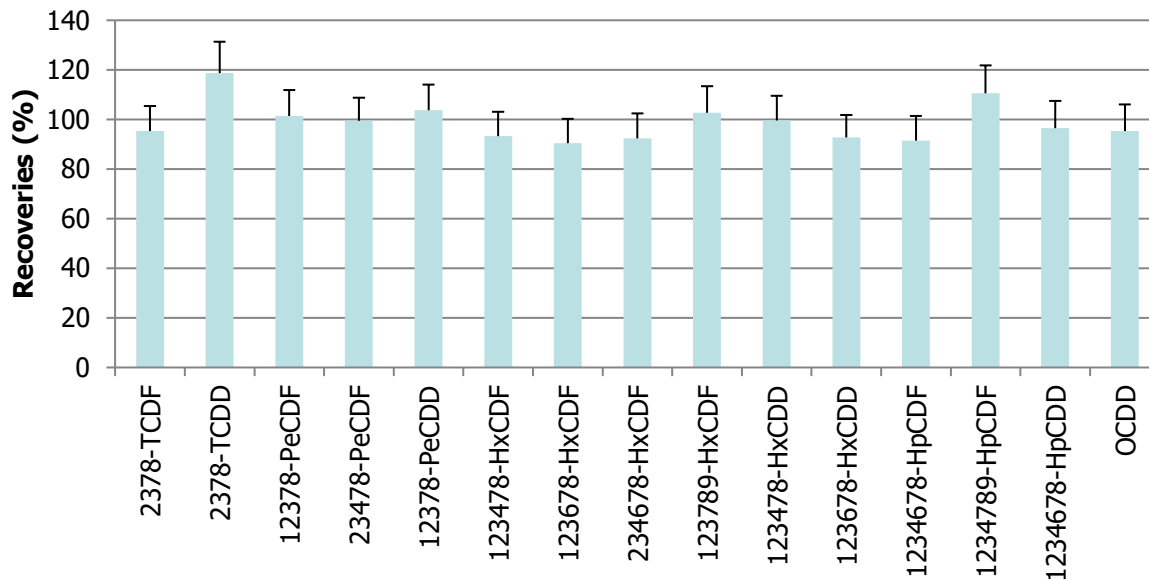


1 L water
SPE ,
transfer to
hexane,
n=6

Extracts in Toluene - PCDD/Fs

- **Stage 1: Connect High Capacity Acid Silica and Alumina (no Carbon) and condition with 60 mL of hexane (vacuum, waste)**
- **Stage 2: Load sample (in 2-10 mL toluene, collect Fraction # 1), rinse loading vials with hexane, elute with 60 mL hexane (collect Fraction # 1), remove acid silica, elute alumina with 30 mL 10% dichloromethane in hexane (collect Fraction # 1)**
- **All 209 PCBs are now in Fraction # 1**
- **Stage 1 Attach carbon column to alumina, elute alumina-carbon with 50 mL dichloromethane (vacuum, waste), all PCDD/Fs now on carbon**
- **Stage 2 Discard alumina, elute carbon in reverse with 50 mL toluene, collect Fraction # 2 with all PCDD/F**

13C recoveries PCDD/Fs fish



10 g fish in
toluene, n=6

Conclusions (1)

- **Samples in toluene (environmental, food): 2-10 mL toluene, separate PCBs and PCDD/Fs completely using hexane and 10% DCM/hexane, followed by DCM and toluene**
- **Reduced hexane volume needed for silica column because of presence toluene**
- **Alternative for samples in toluene: use hexane, DCM and toluene to have mono- and di-ortho PCBs in one fraction, PCDD/F/co-planary PCBs in other fraction**
- **Works also for samples in hexane but more hexane needed in that case for silica elution ("toluene effect" not present)**

Column Kits with various fat removal capacities for samples in hexane

| | | STAGE 1 | | | STAGE 2 | | |
|---------------------|-------------|-------------------|--------------------|-------------------|---------------------|---------------------|------------|
| | | | | | PCBs | Dioxins | |
| | Fat Removal | Hexane | Hexane | Hexane | DCM | Toluene | |
| Column kits | Capacity | conditioning (mL) | sample volume (mL) | Elute Silica (mL) | Alumina-carbon (mL) | Reverse Carbon (mL) | Time (min) |
| Classical Plus | 1.0 g | 20 | 30 | 100 | 50 | 50 | 50 |
| High Capacity | 2.5 g | 40 | 30 | 160 | 50 | 50 | 70 |
| Extra high Capacity | 5.0 g | 60 | 30 | 180 | 50 | 50 | 80 |

Conclusions (2)

- **EZPrep suitable for environmental and food analyses in toluene as solvent. Also suitable for samples in hexane**
- **Can keep PCBs and PCDD/Fs completely separate if so desired. Alternatively have co-planary PCBs in with PCDD/Fs**
- **High sample throughput → 18 samples/hour**
 - **6 samples in parallel per station**
 - **3 stations fit in one hood**
- **System gives excellent recoveries for PCDD/F and PCBs comparable to automated systems**
- **Use of certified pre-packaged columns guarantees low native background**

Conclusions (3)

- **No worries about breakdown or downtime**
- **No washing needed**
- **No cross-contamination**
- **Low cost**



EPH Sample Prep and Analysis

- Soil contamination from diesel fuel, gasoline, heating oil, jet fuel leaks, kerosene or spills is a common occurrence and a global environmental concern.
- EPA 8015B: Total Petroleum Hydrocarbon (TPH) with GC/FID (semi-volatiles)
- Petroleum has > 250 compounds, complex matrix



EPH Sample Prep and Analysis

- Extractable Petroleum Hydrocarbons (EPH): Massachusetts method
- Toxicological approach: evaluate aliphatic and aromatic compounds in extracts
- Semi volatiles evaluated: C₉-C₃₆ aliphatics
- Also range of seventeen aromatics (PAHs)

Features of the MA EPH method (1)

- **Method quantitates aliphatics within two ranges, $C_9 - C_{18}$ and $C_{19} - C_{36}$**
- **PAHs are quantitated within $C_{11} - C_{22}$ range**
- **Collective data reporting**
- **Method can determine health hazards**
- **Also used by other states and some Canadian provinces**



Features of the MA EPH method (2)

- **Uses neutral silica cartridges or columns to separate aliphatics from aromatics in extract**
- **Aliphatics eluted with hexane, aromatics with dichloromethane**
- **Surrogates used to determine quality of separation between aliphatics and aromatics**
- **Breakthrough of naphthalene and 2-methyl naphthalene into aliphatic fraction is regulated**
- **Samples analyzed with GC/FID**

Semi-Automated approach

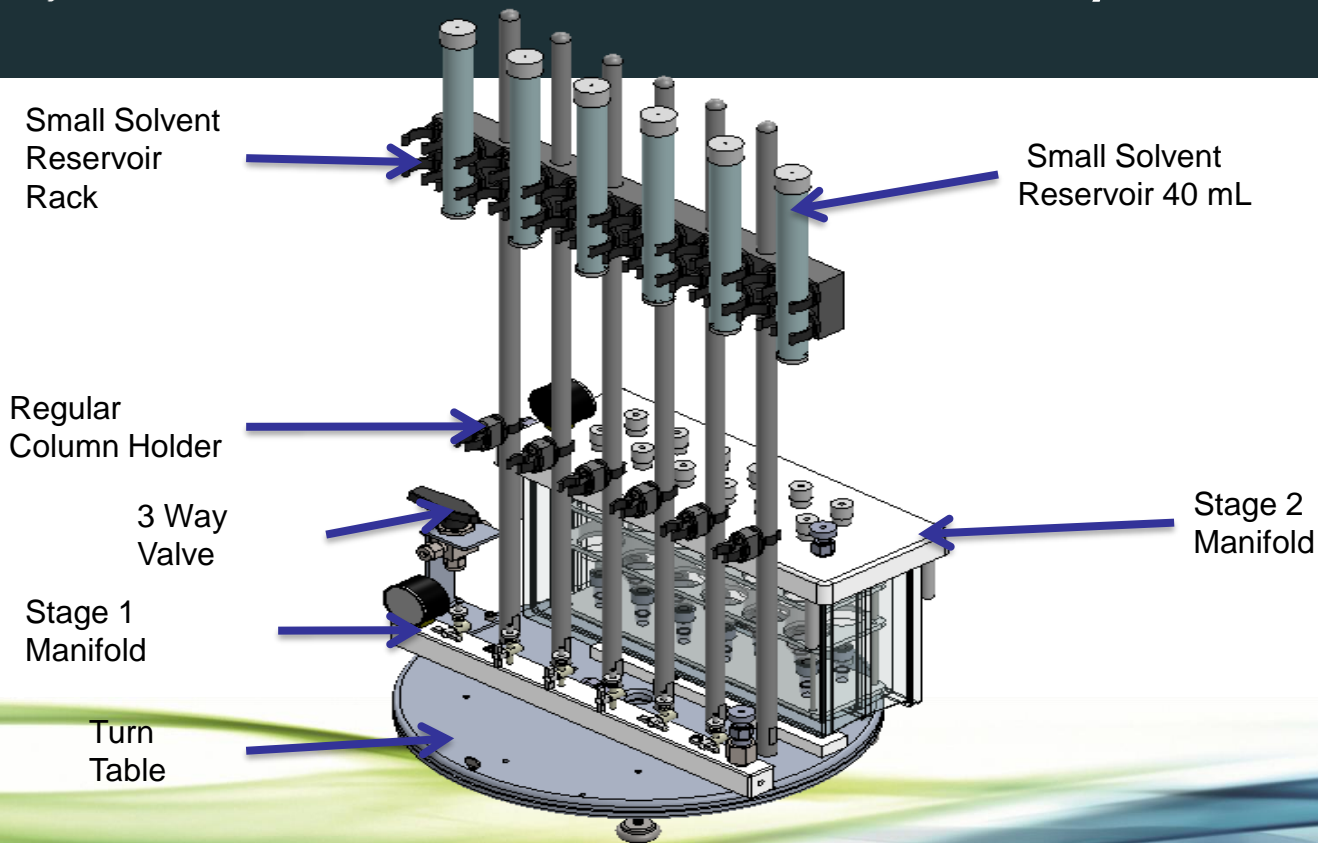
- Manual method is labor intensive, prone to error
- Certified 6 g neutral silica columns can be used with very low native background
- Less interferences in analysis
- Less glassware and solvent use

Semi-Automated System for EPH Analysis

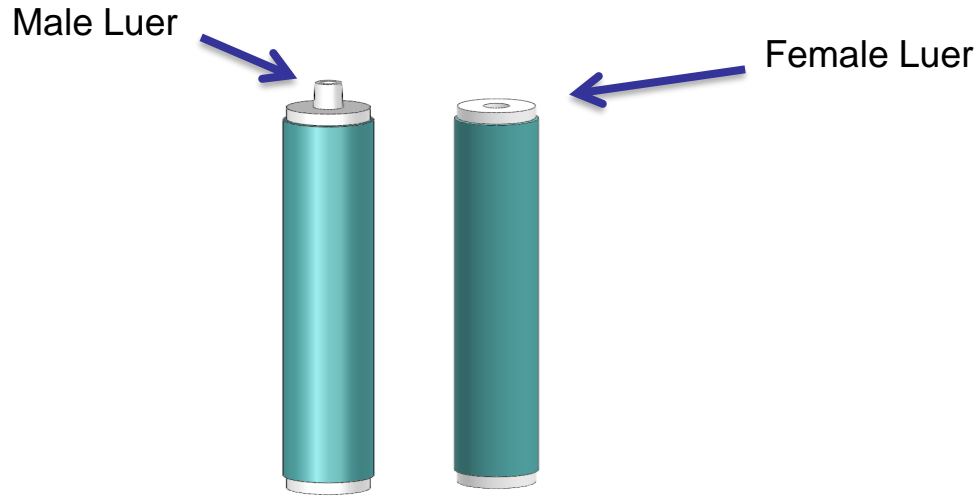
Specification:

- Simple to run, no computerized instrumentation
- Fast: 20 min
- Closed loop system to give a clean background, low level detection
- Use certified columns
- One column per sample
- No capital equipment cost
- No electronics or mechanical equipment to fail
- No downtime

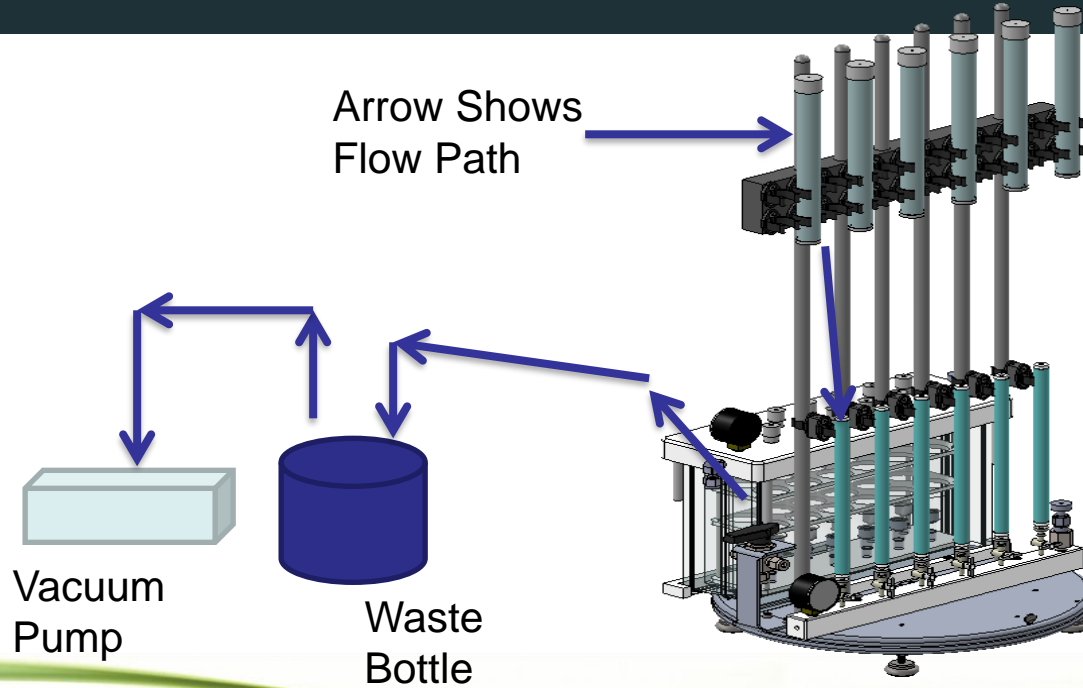
Semi-Automated EZEPH System for EPH



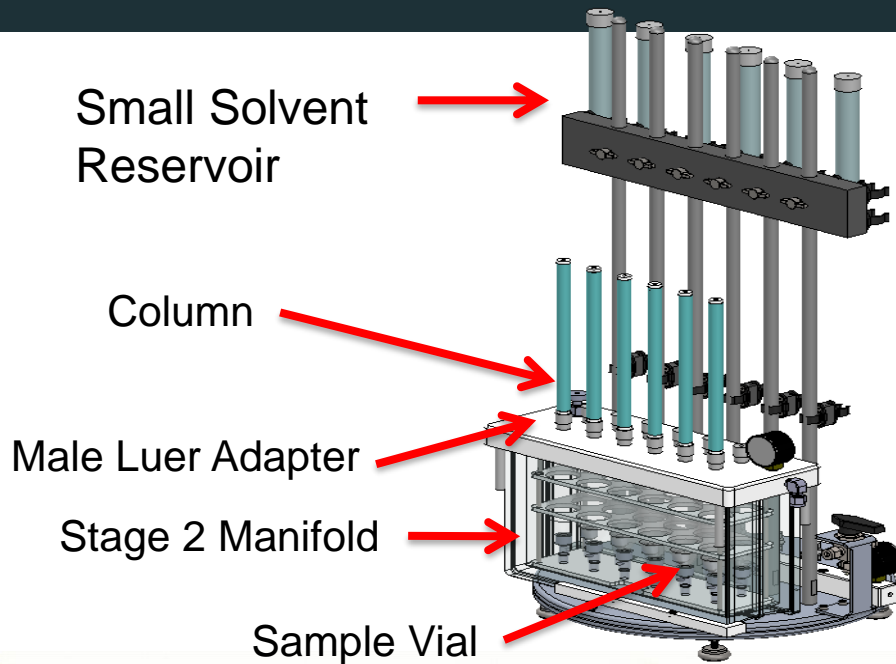
Neutral silica columns



Flow thru system (Stage 1)



Collection (Stage 2)



EZEPH Procedure Stage 1

- **Stage 1:**
- **Assemble silica column with EZPrep set-up**
- **Syringe vial at top is used for conditioning and sample loading**
- **Condition silica column with 30 mL hexane (vacuum, waste)**





EZEPH Procedure Stage 2

- **Stage 2:**
- **Dilute sample extract to 9 mL hexane and spike surrogate compounds (dissolved in 1 mL hexane) into sample extract**
- **Load sample extract onto silica column**
- **Elute column with 10 mL hexane, collecting aliphatic fraction**
- **Elute column with 35 mL dichloromethane, collecting aromatic fraction**



SuperVap 12 50 mLs



Evaporation and Analysis

- **System pre-heated to 30 °C.**
- **Samples evaporated at stable T under 5-6 psi nitrogen.**
- **1 mL extract vial transferred to GC vial (can have direct-to-vial feature)**
- **Analyze on Agilent GC/FID**
- **Samples (hexane) were spiked with 2.5, 12.5 or 25 ug/mL aliphatic and aromatic standards and surrogates before cleanup**



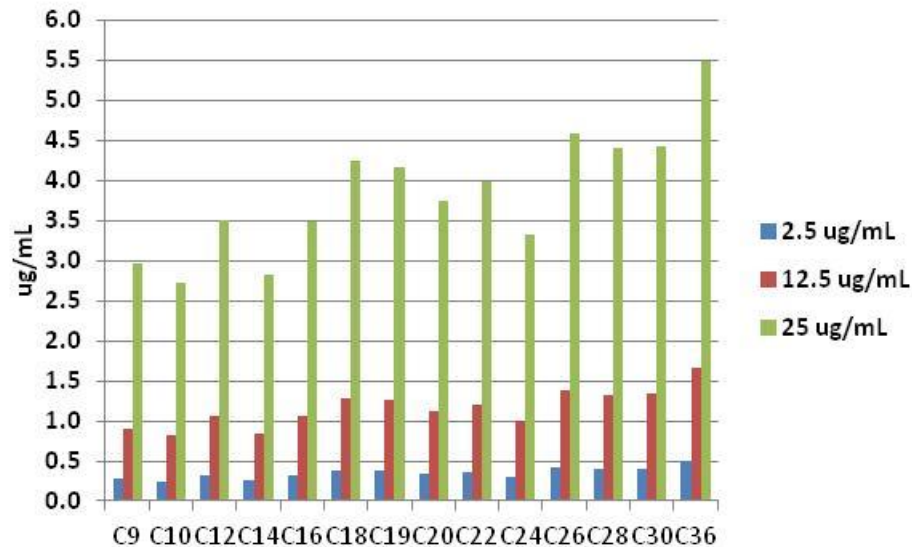
Aliphatic Recoveries (25 ug/mL)

| | Average Recoveries (%) | RSD (%) | EPA Limit (%) |
|-----------------------|---------------------------------------|----------------|------------------------------|
| Nonane (C9) | 74.7 | 7.3 | 30-130 |
| Decane (C10) | 78.6 | 8.4 | 40-140 |
| Dodecane (C12) | 80.9 | 4.5 | 40-140 |
| Tetradecane (C14) | 87.0 | 5.0 | 40-140 |
| Hexadecane (C16) | 81.4 | 3.9 | 40-140 |
| Octadecane (C18) | 85.6 | 3.3 | 40-140 |
| Nonadecane (C19) | 88.6 | 3.5 | 40-140 |
| Eicosane (C20) | 91.5 | 4.1 | 40-140 |
| Docosane (C22) | 92.6 | 4.9 | 40-140 |
| Tetracosane (C24) | 93.2 | 4.9 | 40-140 |
| Hexacosane (C26) | 93.2 | 4.8 | 40-140 |
| Octacosane (C28) | 92.4 | 4.7 | 40-140 |
| Triacontane (C30) | 92.9 | 4.5 | 40-140 |
| Hexatriacontane (C36) | 98.0 | 3.9 | 40-140 |

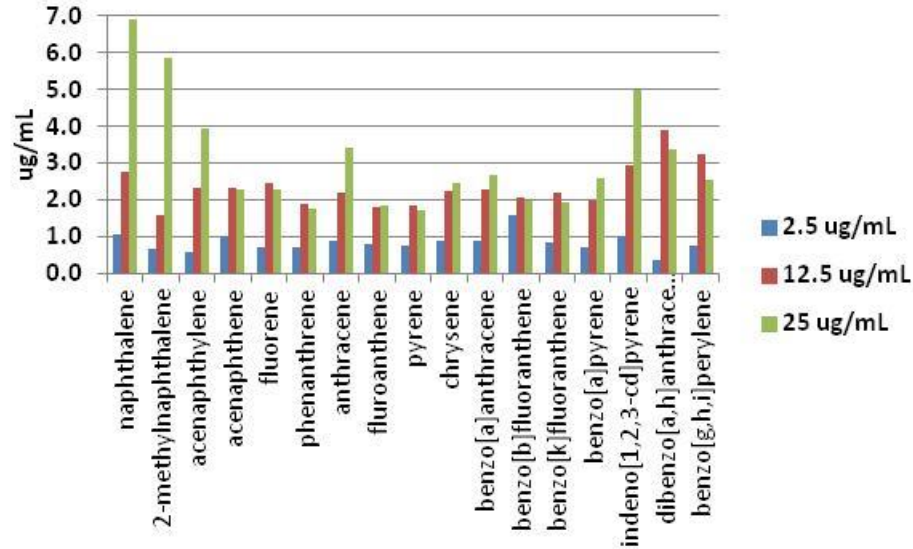
Aromatic Recoveries (25 ug/mL)

| | Average Recoveries (%) | RSD (%) | EPA Limit (%) |
|------------------------|-------------------------------|----------------|----------------------|
| naphthalene | 110.5 | 6.7 | 40-140 |
| 2-methylnaphthalene | 104.2 | 6.3 | 40-140 |
| acenaphthylene | 94.4 | 3.5 | 40-140 |
| acenaphthene | 99.3 | 2.5 | 40-140 |
| fluorene | 107.4 | 1.8 | 40-140 |
| phenanthrene | 109.0 | 1.9 | 40-140 |
| anthracene | 103.1 | 2.4 | 40-140 |
| fluroanthene | 104.8 | 1.8 | 40-140 |
| pyrene | 103.0 | 1.7 | 40-140 |
| chrysene | 97.1 | 2.2 | 40-140 |
| benzo[a]anthracene | 109.6 | 2.9 | 40-140 |
| benzo[b]fluoranthene | 111.9 | 1.9 | 40-140 |
| benzo[k]fluoranthene | 109.0 | 2.3 | 40-140 |
| benzo[a]pyrene | 98.0 | 2.3 | 40-140 |
| indeno[1,2,3-cd]pyrene | 111.6 | 3.1 | 40-140 |
| dibenzo[a,h]anthracene | 96.1 | 2.9 | 40-140 |
| benzo[g,h,i]perylene | 103.7 | 3.5 | 40-140 |

Method Detection Limit Aliphatics



Method Detection Limit Aromatics



Breakthrough

- Average naphthalene breakthrough in aliphatic fraction < 0.02 (limit is < 0.05)
- Average 2-methyl naphthalene breakthrough in aliphatic fraction < 0.01 (limit is < 0.05)



Surrogates (limit 40-140%)

- 1-chloro-octadecane: $83\% \pm 12\%$
- O-terphenyl: $89\% \pm 14\%$
- 2-bromonaphthalene: $70\% \pm 13\%$
- 2-fluorobiphenyl: $104\% \pm 8\%$

Comparison lab X vs EZPrep EPH

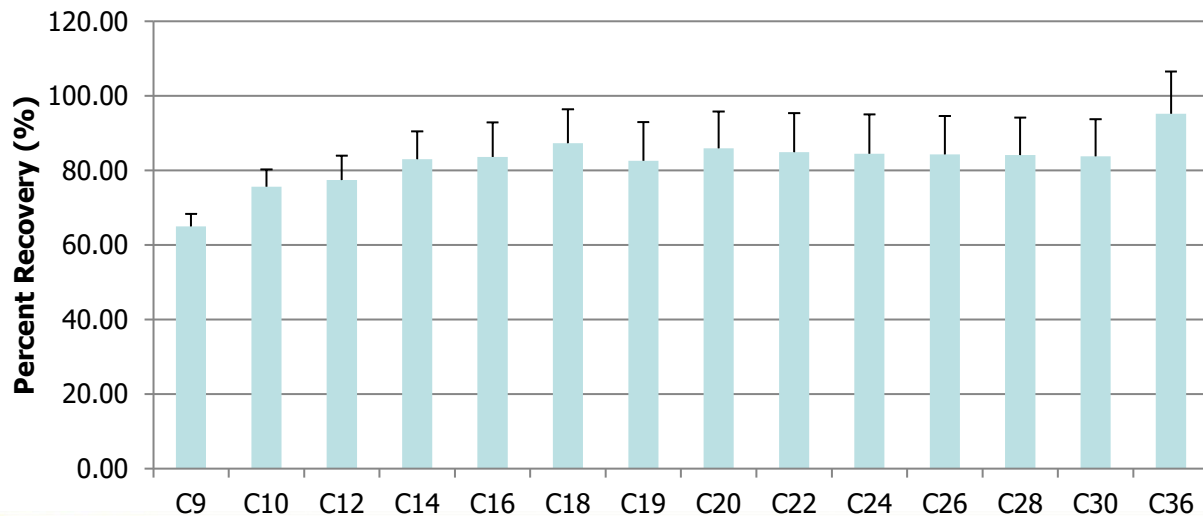
| | C9-C18 Aliphatic | | | C19-C36 Aliphatic | | | C11-C22 Aromatic | |
|-----------|------------------|--------------|--|-------------------|--------------|--|------------------|--------------|
| | Other Lab | Toxic Report | | Other Lab | Toxic Report | | Other Lab | Toxic Report |
| Sample 1 | 17 | 16 | | 144 | 115 | | 191 | 176 |
| Sample 2 | 410 | 292 | | 4314 | 3925 | | 1313 | 1019 |
| Sample 3 | 185 | 136 | | 2335 | 2222 | | 797 | 412 |
| Sample 4 | 33 | 58 | | 57 | 41 | | 88 | 85 |
| Sample 5 | 28 | 46 | | 50 | 68 | | 171 | 163 |
| | | | | | | | | |
| Sample 6 | 82 | 58 | | 188 | 89 | | 140 | 178 |
| Sample 7 | 16 | 30 | | 183 | 171 | | 241 | 226 |
| Sample 8 | 22 | 22 | | 152 | 138 | | 181 | 204 |
| Sample 9 | 27 | 44 | | 119 | 93 | | 213 | 215 |
| Sample 10 | 2931 | 2167 | | 1232 | 1574 | | a | a |
| | | | | | | | | |
| Sample 11 | 171 | 128 | | 89 | 64 | | 113 | 110 |
| Sample 12 | 19 | 20 | | 38 | 40 | | 33 | 73 |
| Sample 13 | 245 | 135 | | 198 | 100 | | 682 | 340 |
| Sample 14 | 61 | 69 | | 364 | 240 | | 334 | 252 |
| Sample 15 | 113 | 91 | | 447 | 200 | | 860 | 740 |
| Sample 16 | 39 | 51 | | 17 | 28 | | 23 | 25 |

**Extracts
from
commercial
lab vs Toxic
Reports lab
Data in
ug/mL**

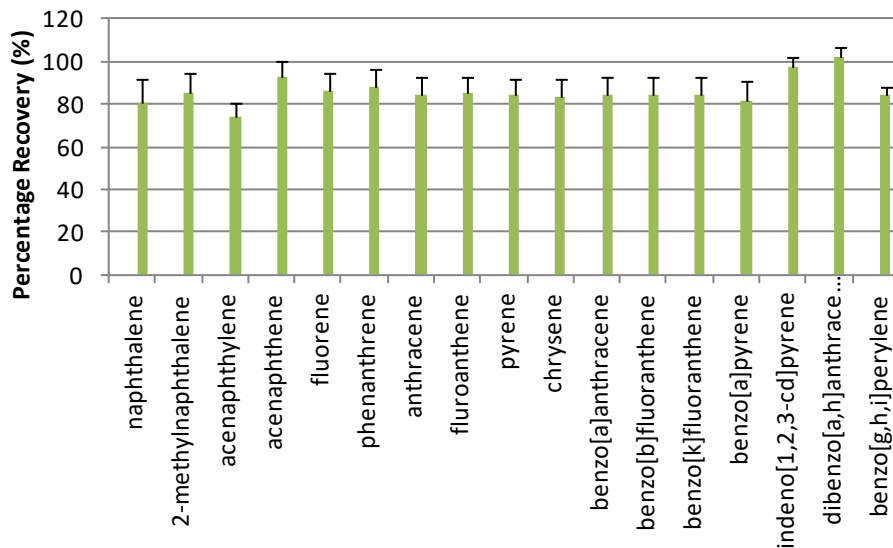
Washington State Method

- **Stage 1: As before but condition 6 g neutral silica column with 10 mL pentane**
- **Stage 2: Spike surrogate compounds in 1 mL pentane or sample extract**
- **Load sample extract onto silica column**
- **Elute column with 15 mL pentane, collecting aliphatic fraction**
- **Elute column with 40 mL dichloromethane, collecting aromatic fraction**

Washington State aliphatics



Washington State aromatics



Conclusions

- Excellent recoveries for aliphatics and aromatics with low RSDs
- All well within MA and WA method windows
- Very good MDL data
- Breakthrough of naphthalene's well within limits
- All surrogates give very good recoveries
- Comparison semi-automated method with manual method for commercial lab samples shows very good agreement
- Can process 6 samples in parallel in 20 min

Questions

